

Plant Archives

Journal homepage: http://www.plantarchives.org DOI Url : https://doi.org/10.51470/PLANTARCHIVES.2025.v25.supplement-1.221

EFFECT OF SELECTION RESPONSE FOR YIELD RELATED TRAITS IN EARLY GENERATION OF FABA BEAN (VICIA FABA L.)

Anjali Kumari Jha¹, Ashish Ranjan^{2*}, Anuj Kumar Choudhary¹, Ajay Bhardwaj³, Raj Bhawan Verma³ and Abhishek Kumar³

¹Department of Horticulture (Vegetable & Floriculture), Bihar Agricultural University, Sabour, Bhagalpur, Bihar, India

²Department of Horticulture, Bhola Paswan Shastri Agricultural College, Purnea, Bihar, India ³Department of Plant Breeding & Genetics, Bhola Paswan Shastri Agricultural College, Purnea, Bihar, India *Corresponding author E-mail: aranjan1978@gmail.com

(Date of Receiving : 22-09-2024; Date of Acceptance : 17-11-2024)

The present experiment was carried out at the research farm of Bhola Paswan Shastri Agricultural College Purnea, Bihar, India, during rabi season of 2022-23 to investigate the selection response for yield related traits in early generations in faba bean. In the current study, genetic interactions were investigated in the F₃ generations of eight faba bean (Vicia faba L.) hybrids. The majority of the traits investigated were positively skewed and controlled by many genes, indicating quantitative inheritance. While in cross combination Bak-1 × Bak-20 non-skewed and platykurtic distribution was observed for ABSTRACT number of seeds per pod at edible stage. The variance, skewness, and kurtosis of all the crosses indicate high level of heterozygosity. All of the crosses outperformed the parents and the national check in terms of producing high-yielding segregants that demonstrated parental diversity. Pod yield per plant and other associated characters, such as number of pods per plant and number of seeds per pod, are controlled by complementary action of multiple genes. Overall, three lines from each cross with superior pod yield were identified, which could be used for varietal development. Keywords: Faba bean, Vicia faba L., skewness, kurtosis, segregation.

Introduction

The faba bean (Vicia faba L.), commonly known as horse bean or broad bean, is a significant minor extensively cultivated in legume crop India. particularly in the North-eastern region. It is predominantly harvested for its dry seeds, which are used as food, while fresh seeds and tender pods are consumed as vegetables. Faba bean is a rich source of essential nutrients, including proteins, carbohydrates, starch, total sugars, vitamins, and minerals (Duranti et al., 2008). Moreover, it contains L-dopa in various parts, a natural compound used in the treatment of Parkinson's disease (Duranti, 2006). However, its consumption is limited by the presence of certain antinutritional factors such as phytic acid, tannins, phenols, trypsin inhibitors, vicine, and covicine, which are associated with hemolytic anemia, a condition

known as favism. Faba bean plays a crucial role in sustainable agriculture by contributing to crop diversity and engaging in biological nitrogen fixation through symbiosis with specific rhizobia bacteria, thereby reducing the need for synthetic fertilizers in cultivation. Although the area under faba bean cultivation in India remains small, it ranks seventh among legume crops globally, following major crops like soybean, groundnut, and common bean.

The global productivity of faba bean remains low, primarily due to the crop's narrow genetic base, which makes it susceptible to various biotic and abiotic stresses (Toker and Mutlu, 2011). Variability is essential for the success of any selection process (Andrade, 2019), as progress in breeding programs relies on the amount of variability present in the population (Vinithashri et al., 2019). To enhance faba bean production, understanding the inheritance patterns of yield and its components is crucial. Genetic variability parameters provide significant value in identifying superior segregants for yield and related traits. Standard measures of variability, such as range, variance, and standard error, are commonly employed to evaluate variability and gene action for desirable traits. Furthermore, information on the frequency distribution of various traits is vital for selecting plants with desirable trait expression (Dinesh *et al.*, 2018). Breeding for higher yield is a primary objective in crop improvement programs (Woyann *et al.*, 2019).

Since yield is polygenically controlled and highly influenced by environmental factors (Santos *et al.*, 2014), selection based solely on yield may not be effective. Therefore, yield improvement can be achieved through indirect selection of component traits (Luz *et al.*, 2011) that are highly heritable and strongly correlated with yield. The distribution properties of skewness and kurtosis provide insights into the nature of gene action and the number of genes controlling each trait, respectively. Consequently, the present study aimed to examine the genetics of different traits in association with yield and its component traits, utilizing third and fourth-degree statistics in the early generations of eight different faba bean crosses.

Material and Methods

The research was conducted at the experimental farm of Bhola Paswan Shastri Agricultural College, Purnea, Bihar, India. The study material comprised F_3 generations from eight faba bean crosses, specifically Bak-1 × Bak-5, Bak-1 × Bak-11, Bak-1 × Bak-20, Bak-2 × Bak-5, Bak-2 × Bak-11, Bak-2 × Bak-20, Bak-12 × Bak-5, and Bak-16 × Bak-20, along with the parent lines and a national check variety, Vikrant. The plants were cultivated under field conditions during the *rabi* season of 2022-23, adhering to standard crop

management practices to ensure healthy growth. Seeds were sown in the first week of November, with a spacing of 45 x 15 cm between plants. A total of 640 plants were selected for the study, with 80 plants from each cross. Observations were made on 13 quantitative traits, including days to flowering, days to first pod picking, plant height (cm), number of branches, internodal length, node of the first flower, pod length (cm), pod diameter (cm), number of fruit clusters per plant, number of pods per plant, pod yield per plant (g), number of seeds per pod, and harvesting span. Data were recorded from individual F₃ plants and from five randomly selected plants of each parent and the national check. The range and variance of yield and associated traits were determined using an analysis toolkit in Microsoft Excel. Skewness (K3) and thirddegree statistics, along with kurtosis (K4) and fourthdegree statistics, were calculated using Minitab software to analyze the nature of trait distributions.

Additionally, the yield of the 640 F_3 progenies was compared to that of the best check cultivar, Vikrant, which was grown in the field alongside the test lines. F_3 progenies exhibiting higher pod yield per plant than their respective parents and the check cultivar were selected for further evaluation. The standard error of the mean (SE), standard deviation, and variance were analyzed using the formula provided by Jasani *et al.* (2018).

Results and Discussion

The range demonstrated considerable variability both among and within the F3 populations for all thirteen quantitative traits across the crosses (Table 1). This variability can be attributed to the broad differences among F2 plants and the presence of heterozygosity at several loci where the parents exhibited genetic differences.

Table 1: Range of thirteen traits in eight populations of faba bean

8	Bak-1 ×	Bak-1 ×	Bak-1 ×	$Bak-2 \times$	$Bak-2 \times$	Bak-2 \times	Bak-12 ×	Bak-16 x
Genotypes	Bak-5	Bak-11	Bak-20	Bak-5	Bak-11	Bak-20	Bak-5	Bak-20
Traits					Range			
PH	22-57	28-60	23-66	31-79	33-71	29-68	42-77	40-88
NB	2-13	2-23	2-10	2-10	2-12	2-13	2-9	2-12
IL	2.70-4.40	2.30-4.40	2.80-4.60	2.00-6.50	2.20-6.00	2.00-4.30	1.80-4.30	2.00-4.80
NFF	1-5	1-5	1-6	2-8	2-6	2-6	1-5	1-9
DFF	60-101	56-87	52-87	44-108	47-86	46-88	43-78	47-88
DPP	99-115	96-114	99-112	99-115	89-115	94-117	92-112	103-114
NFCP	0-10	0-11	0-11	0-14	0-9	0-8	0-14	0-20
NPP	10-55	5-75	6-62	6-77	8-48	6-55	10-55	6-61
PL	4.82-6.20	4.14-5.92	4.26-6.08	3.37-6.39	4.22-6.86	4.45-5.95	4.20-6.00	3.98-7.18

PD	0.87-1.18	0.86-1.12	0.88-1.03	0.89-1.04	0.84-1.08	0.84-1.04	0.87-1.06	0.89-1.24
HS	4-17	2-13	2-13	3-19	2-16	2-10	2-18	4-17
NSPPES	1.40-3.60	1.40-3.60	1.60-3.20	1.80-3.40	1.60-3.20	1.80-3.80	1.80-3.20	2.20-3.80
РҮ	26.97-	10.12-171	16.93-	18.90-	12.60-	13.50-	14.40-	17.36-
F I	129.29	10.12-171	125.05	130.40	120.96	104.40	109.33	135.12

PH: Plant height (cm), NB: No. of branches per plant, IL: Internodal length (cm), NFF: Node of first flower, DFF: Days to first flowering, DPP: Days to first pod picking, NFCP: No. of fruit cluster per plant, NPP: No. of pods per plant, PL: Green pod length (cm), PD: Pod diameter (cm), HS: Harvesting span, NSPPES: No. of seeds per pod at edible stage, PY: Green pod yield per plant (g)

Skewness and kurtosis in F_3 population of Bak-1 \times Bak-5

The range of all the traits revealed a highly significant difference for key characteristics such as the number of branches per plant, plant height, number of pods per plant, and pod yield per plant across the crosses, indicating substantial variability in all the studied traits. The analysis of trait distribution using skewness and kurtosis provides valuable insights into the nature of gene action and the number of genes controlling the traits (Govintharaj et al., 2018). Positive skewness is typically linked to complementary gene action, while negative skewness indicates duplicate (additive × additive) gene interactions. Kurtosis tends to be negative or close to zero (platykurtic) in the absence of gene interaction but becomes positive (leptokurtic) when gene interaction is present (Ramadhan et al., 2018). Understanding gene interactions is essential for improving selection efficiency in breeding programs.

Positive skewness combined with a platykurtic distribution suggested the involvement of dominancebased complementary gene interaction and a large number of genes with decreasing effects in the inheritance of traits such as the number of branches per plant, internodal length (cm), node of the first flower, number of pods per plant, pod length (cm), pod diameter (cm), harvesting span, number of seeds per pod at the edible stage, and green pod yield per plant (g) in the cross Bak-1 \times Bak-5. Similar results were observed for traits such as plant height (cm), number of branches per plant, node of the first flower, days to first flowering, pod diameter (cm), harvesting span, and green pod vield per plant (g) in the cross Bak-1 \times Bak-11, and for traits like the number of branches per plant, node of the first flower, number of pods per plant, pod length (cm), harvesting span, number of seeds per pod at the edible stage, and green pod yield per plant (g) in Bak-1 \times Bak-20. This pattern continued with traits such as the number of branches per plant, internodal length (cm), node of the first flower, days to first flowering, days to first pod picking, number of pods per plant, harvesting span, number of seeds per

pod at the edible stage, and green pod yield per plant (g) in the cross Bak-2 \times Bak-5, and for plant height (cm), number of branches per plant, internodal length (cm), node of the first flower, number of pods per plant, pod diameter (cm), harvesting span, number of seeds per pod at the edible stage, and green pod yield per plant (g) in Bak-2 × Bak-11. Furthermore, internodal length (cm), node of the first flower, days to first flowering, number of fruit clusters per plant, number of pods per plant, pod length (cm), pod diameter (cm), number of seeds per pod at the edible stage, and harvesting span in Bak-2 × Bak-20, and traits such as plant height (cm), number of branches per plant, internodal length (cm), node of the first flower, days to first flowering, days to first pod picking, number of pods per plant, number of seeds per pod at the edible stage, and harvesting span in Bak-12 × Bak-5 showed similar inheritance patterns. The same was observed for plant height (cm), node of the first flower, days to first flowering, number of pods per plant, and green pod yield per plant (g) in the cross Bak-16 \times Bak-20. These traits exhibit dominant and dominance-based complementary epistasis in their inheritance, suggesting that intensive selection is necessary to achieve significant genetic improvements in these traits.

On the other hand, positive skewness and leptokurtic distribution were found for traits such as the number of branches per plant, number of fruit clusters per plant, number of pods per plant, and green pod yield per plant (g) in Bak-1 × Bak-11, number of fruit clusters per plant in Bak-1 × Bak-20, number of fruit clusters per plant in Bak-2 × Bak-5, number of fruit clusters per plant and pod length (cm) in Bak-2 × Bak-11, the number of branches per plant in Bak-2 × Bak-20, the number of seeds per pod at the edible stage in Bak-12 × Bak-5, and the number of branches per plant and number of fruit clusters per plant in Bak-2 × Bak-20, indicating the presence of complementary gene interactions. These traits are governed by a few dominant genes, most of which have increasing effects.

Negative skewness and a platykurtic distribution were observed for traits like plant height (cm), days to

first flowering, and days to first pod picking in Bak-1 × Bak-5, and for internodal length (cm), days to first pod picking, pod length (cm), and the number of seeds per pod at the edible stage in Bak-1 \times Bak-11. In Bak-1 \times Bak-20, traits such as plant height (cm), internodal length (cm), days to first flowering, days to first pod picking, and pod diameter (cm) showed negative skewness. Similarly, for Bak-2 \times Bak-5, plant height (cm), pod length (cm), and pod diameter (cm) exhibited negative skewness, while days to first flowering and days to first pod picking showed this pattern in Bak-2 × Bak-11. Traits like plant height (cm) and days to first pod picking in Bak-2 × Bak-20, pod length (cm), pod diameter (cm), and the number of seeds per pod at the edible stage in Bak-12 \times Bak-5, and days to first pod picking, pod length (cm), pod diameter (cm), harvesting span, and the number of seeds per pod at the edible stage in Bak-16 \times Bak-20, suggested the involvement of a large number of genes,

most of which exhibited duplicate inheritance and dominance with increasing effects. Under mild selection, utilizing the existing variability, rapid genetic gains can be achieved for all these traits exhibiting duplicate gene action.

The number of seeds per pod at the edible stage in the cross combination Bak-1 × Bak-20 exhibited a nonskewed and platykurtic distribution, indicating that the inheritance of this trait is governed by a few segregating genes, with most showing a decreasing effect and dominance-based complete interaction. To achieve maximum genetic gain for these traits, intense selection would be necessary. Similar findings were previously reported by Chandrakant *et al.* (2015) in dolichos bean, Prabhu *et al.* (2015), Ajay *et al.* (2016) in pigeon pea, Sumathi *et al.* (2018) in maize, Mariyammal *et al.* (2019) in green gram, Kamdar *et al.* (2020) in groundnut, and Kanavi *et al.* (2020) in green gram.

Table 2: Skewness and kurtosis in F₃ Population of Bak-1 × Bak-5 and Bak-1 × Bak-11

				F ₃ Population	l				
		Bak-1	× Bak-5		Bak-1 × Bak-11				
S.No.	Traits	Skewness	Kurtosis	Variance	Skewness	Kurtosis	Variance		
1.	PH	-0.05	-0.12	46.46	0.25	-0.62	62.50		
2.	NB	0.66	-0.09	6.79	2.82	12.19	10.53		
3.	IL	0.07	-0.53	0.16	-0.13	-1.03	0.30		
4.	NFF	0.53	-0.68	1.11	0.76	0.10	1.22		
5.	DFF	-1.13	1.69	64.00	0.28	-0.82	78.42		
6.	DPP	-0.18	-0.48	14.52	-0.09	-0.26	13.47		
7.	NFCP	1.15	1.31	4.85	1.96	5.24	4.15		
8.	NPP	0.48	0.38	81.19	1.84	3.97	182.04		
9.	PL	0.22	-0.44	0.11	-0.53	0.57	0.13		
10.	PD	0.61	1.05	0.00	0.84	0.61	0.00		
11.	HS	0.06	-0.73	9.12	0.62	-0.38	7.54		
12.	NSPPES	0.01	-0.11	0.17	0.31	0.52	0.18		
13.	PY	0.14	-0.40	555.43	1.96	4.54	932.98		

Skewness: positive (+) and negative (-), Kurtosis value less than 3 indicates platykurtic, more than 3 leptokurtic and three indicates mesokurtic

Table 3: Skewness and kurtosis in F₃ Population of Bak-1 × Bak-20 and Bak-2 × Bak-5

			I	F ₃ Population						
		Bak-1 × Ba	k-20			Bak-2 × Bak-5				
S.No.	Traits	Skewness	Kurtosis	Variance	Skewness	Kurtosis	Variance			
1.	PH	-0.05	-0.35	84.33	-0.13	-0.47	91.58			
2.	NB	0.73	-0.19	4.51	0.42	-0.22	3.48			
3.	IL	-0.19	-0.90	0.26	0.79	0.18	0.91			
4.	NFF	0.49	0.27	1.27	0.91	0.49	1.75			
5.	DFF	-0.10	-0.78	64.92	0.29	0.69	145.43			
6.	DPP	-0.24	-1.07	14.27	0.22	-0.60	17.10			
7.	NFCP	2.87	13.43	2.59	1.81	5.91	5.57			
8.	NPP	1.09	1.29	130.59	1.15	1.74	193.65			
9.	PL	0.46	2.77	0.07	-0.87	1.03	0.37			
10.	PD	-0.46	0.40	0.00	-0.28	-0.46	0.00			
11.	HS	0.33	-0.10	5.34	0.04	-0.72	19.22			
12.	NSPPES	0.00	-0.77	0.13	1.15	1.03	0.13			
13.	PY	0.99	0.86	531.24	1.09	1.48	525.36			

				F ₃ Population					
		Bak-2 × Ba	ak-11		Bak-2 × Bak-20				
S.No.	Traits	Skewness	Kurtosis	Variance	Skewness	Kurtosis	Variance		
1.	PH	0.19	0.08	57.69	-0.65	0.72	45.69		
2.	NB	1.36	2.99	3.62	1.67	5.88	3.17		
3.	IL	1.13	1.71	0.50	0.58	0.35	0.23		
4.	NFF	1.02	0.99	0.92	0.98	2.08	0.59		
5.	DFF	-0.07	-0.04	75.18	0.05	-0.51	112.97		
6.	DPP	-0.51	0.34	27.17	-0.62	-0.17	27.87		
7.	NFCP	2.02	5.93	2.79	1.32	1.32	3.58		
8.	NPP	1.20	1.46	76.42	1.18	0.74	141.39		
9.	PL	1.26	3.92	0.19	0.41	-0.39	0.11		
10.	PD	0.50	-0.10	0.00	0.35	-1.16	0.00		
11.	HS	1.21	1.57	8.17	1.14	2.12	2.59		
12.	NSPPES	0.02	0.05	0.09	1.71	4.91	0.11		
13.	PY	1.14	1.78	408.77	0.87	-0.09	516.02		

Table 4: Skewness and kurtosis in F_3 Population of Bak-2 × Bak-11 and Bak-2 × Bak-20

Table 5: Skewness and kurtosis in F₃ Population of Bak-1 2× Bak-5 and Bak-16× Bak-20

			F	3 Population						
		Bak-12 × B	Bak-5		Bak-16 × Bak-20					
S.No.	Traits	Skewness	Kurtosis	Variance	Skewness	Kurtosis	Variance			
1.	PH	0.64	1.25	36.67	0.38	0.18	91.29			
2.	NB	0.49	-0.09	3.45	0.77	-0.08	6.60			
3.	IL	0.39	-0.55	0.37	0.36	-0.44	0.34			
4.	NFF	0.40	-0.24	0.92	1.50	2.30	2.65			
5.	DFF	0.69	-0.18	68.22	0.67	-0.30	100.71			
6.	DPP	0.11	-0.67	21.98	-0.49	0.13	4.74			
7.	NFCP	2.47	9.94	4.79	2.73	10.15	11.42			
8.	NPP	0.93	0.52	93.79	0.13	-0.54	157.57			
9.	PL	-0.03	-0.48	0.17	-0.18	0.32	0.33			
10.	PD	-0.52	-0.03	0.00	-0.22	0.16	0.00			
11.	HS	0.75	0.04	15.26	-0.45	0.50	6.27			
12.	NSPPES	0.38	-0.46	0.12	1.49	2.70	0.10			
13.	РҮ	0.69	0.41	384.54	0.04	-0.94	960.66			

Selection of superior lines

All the 640 F_3 progenies were classified to identify superior lines with higher pod yield per plant when compared with the yield of parents and check cultivars. On the basis of yield best three lines were selected among 80 plants from each cross. Considering all the important and yield attributing traits, on which selection is based, best three segregants in the cross combination Bak-1 \times Bak-5, *viz.* plant numbers 38, 54 and 36 that produced pods more than 120 g per plant with greater than 50 pods per plant along with more than 3.00 number of seeds per pod were identified as a best segregants.

Table 6: Selected segregants on the basis of yield attributing traits in Bak-1 × Bak-5 and Bak-1 × Bak-11

Bak-1 × Bak-5			Segregants			Bak-1 × Bak-11		Segregants		
Traits	Parents	Check	Plant	Plant	Plant	Parents	Check	Plant	Plant	Plant
TTaits	mean	mean	No. 38	No. 54	No. 36	mean	mean	No. 1	No. 28	No. 57
PY	76.39	68.85	129.29	127.35	124.97	77.65	68.85	171.00	152.76	131.80
NPP	39.00	37.92	55	51	50	40.00	37.92	75	67	57
NSPPES	2.30	2.92	3.60	3.30	3.20	2.31	2.92	3.00	2.60	2.40
NFCP	2.70	4.89	10	9	3	2.15	4.89	7	11	5

Bak-1 × Bak-20			Segregants			Bak-2 × Bak-5		Segregants		
Traits	Parents	Check	Plant	Plant	Plant	Parents	Check	Plant	Plant	Plant
TTalts	mean	mean	No. 32	No. 1	No. 11	mean	mean	No. 16	No. 14	No. 13
PY	74.65	68.85	125.05	115.18	105.35	66.25	68.85	130.40	126.40	117.80
NPP	39.50	37.92	62	53	52	33.50	37.92	77	68	61
NSPPES	2.28	2.92	2.20	3.20	2.80	2.26	2.92	3.40	3.30	3.00
NFCP	2.65	4.89	11	7	5	1.90	4.89	6	4	8

Table 7: Selected segregants on the basis of yield attributing traits in Bak-1 × Bak-20 and Bak-2 × Bak-5

Table 8: Selected segregants on the basis of yield attributing traits in Bak-2 × Bak-11 and Bak-2 × Bak-20

Bal	$Bak-2 \times Bak-11$			Segregants			Bak-2 \times Bak-20		Segregants		
Traits	Parents mean	Check mean	Plant No. 37	Plant No. 1	Plant No. 38	Parents mean	Check mean	Plant No. 24	Plant No. 32	Plant No. 30	
PY	67.51	68.85	120.96	93.30	93.24	64.51	68.85	104.40	102.60	100.80	
NPP	34.50	37.92	48	37	37	34.00	37.92	55	54	53	
NSPPES	2.27	2.92	2.80	2.00	2.00	2.24	2.92	3.40	3.20	3.80	
NFCP	1.35	4.89	8	3	5	1.85	4.89	8	4	6	

Table 9: Selected segregants on the basis of yield attributing traits in Bak-12 \times Bak-5 and Bak-16 \times Bak-20

Bal	Bak-12 × Bak-5			Segregants			Bak-16 × Bak-20		Segregants		
Traits	Parents mean	Check mean	Plant No. 1	Plant No. 65	Plant No. 35	Parents mean	Check mean	Plant No. 53	Plant No. 74	Plant No. 51	
PY	78.73	68.85	109.33	98.10	97.74	86.12	68.85	135.12	130.88	129.12	
NPP	47.50	37.92	55	50	49	49.50	37.92	61	58	57	
NSPPES	2.33	2.92	3.20	2.80	2.60	2.63	2.92	3.20	3.80	3.40	
NFCP	2.30	4.89	7	5	3	2.75	4.89	11	6	3	

In the cross combination Bak-1 \times Bak-11, plant numbers 1, 28, and 57 were identified as superior, producing over 130 g per plant, with more than 55 pods per plant, over 2.20 seeds per pod. Similarly, in the Bak-1 \times Bak-20 cross combination, plant numbers 32, 1, and 11 were identified as the best segregants, yielding over 100 g per plant with more than 50 pods, over 2.20 seeds per pod. In the cross combination Bak- $2 \times \text{Bak-5}$, plant numbers 16, 14, and 13 were identified for producing over 115 g per plant, with more than 60 pods, over 3.00 seeds per pod. For the Bak- $2 \times$ Bak-11 combination, plant numbers 37, 1, and 38 were identified for producing more than 90 g per plant, with over 35 pods, more than 1.90 seeds per pod. In the cross combination Bak-2 × Bak-20, plant numbers 24, 32, and 30 were recognized as the best segregants, producing more than 100 g per plant with over 50 pods, more than 3.00 seeds per pod. In the Bak-12 × Bak-5 cross, plant numbers 1, 65, and 35 were identified for producing over 97 g per plant, with more than 48 pods, over 2.50 seeds per pod. Similarly, in the Bak-16 × Bak-20 cross combination, plant numbers 53, 74, and 51 were recognized for producing over 125 g per plant, with more than 60 pods, over 3.00 seeds per pod. These lines will undergo further

testing to confirm their yield stability over time and their potential for future use. Previous studies in chickpea have demonstrated that superior lines derived from transgressive segregation exhibit higher progeny values compared to their parents (Koseoglu et al., 2017; Ceylan et al., 2019). Genetic research suggests that transgressive segregation typically arises from the combination of alleles from both parents, with complementary gene effects distributed among them (Risenberg et al., 1999). In F3 generations, transgressive segregants may result from dominance × dominance and additive × additive interactions, which are fixable due to the recombination of genes with positive effects. This phenomenon has been previously observed in lentils for seed yield per plant (Chahota et al., 2007), peanuts for mature pod weight and grain weight per plant (Shreya et al., 2017), rice for flowering days (Koide et al., 2019), and pigeon pea for earliness (Srivastava & Saxena, 2019).

Conclusion

The process of hybridization and recombination generated substantial genetic variability in the faba bean F_3 populations analyzed. Rare gene combinations in segregating generations, accompanied by additive

gene action, resulted in extreme phenotypes that surpassed the values of the parent plants. A high degree of variability in traits was observed in the F3 generation, attributed to heterozygosity. The majority of the traits studied exhibited positive skewness and were controlled by multiple genes, indicating quantitative inheritance. Based on yield performance in comparison to the parents and the check cultivar, a total of twenty-four superior progenies were identified within the F3 population, each demonstrating higher pod yield per plant than both the parents and the respective check cultivars.

References

- Ajay, B.C., Byregowda, M., Veerakumar, G.N., Ganapathy, K.N., Meena, M., Babu, H.P. and Reena, M. (2016). Genetic association and frequency distribution in segregating generations derived from pigeon pea crosses. *The Indian Journal of Genetics and Plant Breeding*, **76**,181-186.
- Andrade, J.A.D.C. (2019). Genetic variability and breeding potential of Flintisa Composite of maize in two levels of technology. *Crop Breeding and Applied Biotechnology*, 19, 145-152.
- Ceylan, F.O., Adak, A., Sari, D., Sari, H. and Toker, C. (2019). Unveiling of suppressed genes in interspecific and backcross populations derived from mutants of Cicer species. *Crop and Pasture Science*, **70**, 254-262.
- Chahota, R.K., Kishore, N., Dhiman, K. C., Sharma, T.R. and Sharma, S.K. (2007). Predicting transgressive segregants in early generation using single seed descent methodderived micro-macrosperma genepool of lentil (*Lens culinaris* Medikus). *Euphytica*, **156**, 305-310.
- Chandrakant, Ramesh, S., Vaijayanthi, P.V., Gowda, M.B., Rao, M.A., Keerthi, C.M. and Shivakumar, M.S. (2015). Impact of F2bi-parental on quantitative traits inter relationships and frequency of transgressive segregants in Dolichos bean (*Lablab purpureus* L.) *Electronic Journal* of Plant Breeding, 6(3), 723-728.
- Dinesh, H.B., Viswanatha, K.P., Lohithaswa, H.C., Pavan, R. and Poonam, S. (2018). Genetic association estimates using third and fourth degree statistics in early segregating generations of cowpea. *International Journal of Current Microbiology and Applied Sciences*, 7, 867-873.
- Duranti, M., Consonni, A., Magni, C., Sessa, F. and Scarafoni, A. (2008). The major proteins of lupin seed: characterisation and molecular properties for use as functional and nutraceutical ingredients. *Trends in Food Science and Technology*, **19**(12), 624-633.
- Duranti, M. (2006). Grain legume proteins and nutraceutical properties. *Fitoterapia*, 77(2), 67-82. 8.
- Govintharaj, P., Manonmani, S. and Robin, S. (2018). Variability and genetic diversity study in an advanced segregating population of rice with bacterial blight resistance genes introgressed. *Ciência e Agrotecnologia*, 42, 291-296.
- Koide, Y., Sakaguchi, S., Uchiyama, T., OTA, Y., Tezuka, A., Nagano, A.J., Ishiguro, S., Takamure, I. and Kishima, Y. (2019). Genetic properties responsible for the transgressive segregation of days to heading in rice. *G3: Genes, Genomes, and Genetics*, 9, 1655-1662.

- Koseoglu, K., Adak, A., Sari, D., Sari, H., Ceylan, F.O. and Toker, C. (2017). Transgressive segregations for yield criteria in reciprocal interspecific crosses between *Cicer arietinum* L. and *C. reticulatum* Ladiz. *Euphytica*, 213, 116.
- Luz, L.N.D., Santos, R.C.D. and Melo, Filho, P.D.A. (2011). Correlations and path analysis of peanut traits associated with the peg. *Crop Breeding and Applied Biotechnology*, **11**, 88-95.
- Mariyammal, M., Pandiyan, C., Vanniarajan, J.S., Kennedy, N. and Senthil. (2019). Studies in three F₂ populations of Grren gram (Vigna radiata L. Wilzeck). *International Journal of Genetics*, **11**(2), 564-566.
- Prabhu, R., Manivannan, N., Mothilal, A. and Ibrahim, S.M. (2015). Nature and degree of distribution for yield and yield attributes in six backcross populations of groundnut (*Arachis hypogaea* L.). *Plant Archives*, **15**, 997-1001.
- Ramadhan, F., Suwarno, W.B., Nindita, A. and Aswidinnoor, H. (2018). Genetic analysis of panicle architecture using two F2 population in Rice. *Indonesian Journal of Agronomy*, **46**, 1-8.
- Risenberg, L.H., Archer, M.A. and Wayne, R.K. (1999). Transgressive segregation, adaptation and speciation. *Heredity*, 83, 363-372.
- Santos, A.D., Ceccon, G., Davide, L.M.C., Correa, A.M. and Alves, V.B. (2014). Correlations and path analysis of yield components in cowpea. *Crop Breeding and Applied Biotechnology*, 14, 82-87.
- Showkath, B.M., Jagadeesh, B.N., Ramesh, S., Keerthi, C.M. and Sowmya, H.H. (2017). Third and Fourth Degree Statistics-Based Genetics of Quantitative Traits in Dolichos Bean (*Lablab purpureus* L.). *International Journal of Currrent Microbiology and Applied Science*, 6(10), 2551-2558.
- Shreya, Ainmisha, and Vashanti, RP. (2017). Transgressive segregation study in F₃ population of four groundnut crosses. *International Journal of Current Microbiology and Applied Sciences*, **6**, 2054-2059.
- Srivastava, R.K. and Saxena, K.B. (2019). The earliest maturing pigeonpea [*Cajanus cajan* (L.) Millspaugh] germplasm bred at ICRISAT. *Genetic Resources and Crop Evolution*, **6**, 763-766.
- Sumathi, K., Natesan, S. and Ganesan, K.N. (2018). Studies on Skewness and Kurtosis of Sorghum Downy Mildew Resistant BC3F3 Progenies in Maize. *International Journal of Current Microbiology and Applied Sciences*, 7(6), 3834-3838.
- Toker, C. and Mutlu, N. (2011). Breeding for abiotic stresses. In Pratap A and Kumar J (eds) Biology and breeding of food legumes. *CAB International*, Walingford, p. 241-261.
- Vinithashri, G., Manivannan, N., Viswanathan, P.L. and Selvakumar, T. (2019). Genetic variability, heritability and genetic advance of yield and related traits in F₃ generation of groundnut (*Arachis hypogaea* L.). *Electronic Journal of Plant Breeding*, **10**, 1292-1297.
- Woyann, L.G., Zdziarski, A.D., Baretta, D., Meira, D., Dallacorte, L.V. and Benin, G. (2019). Selection of highyielding, adapted and stable wheat lines in preliminary trials. *Crop Breeding and Applied Biotechnology*, **19**, 412-419.